OXOID QUALITY ASSURANCE PRODUCT SPECIFICATION

RAPPAPORT-VASSILIADIS SOYA PEPTONE (RVS) BROTH CM0866

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Typical Formula*

Soya peptone	grams per litre	4.5
Sodium chloride		7.2
Potassium dihydrogen phosphate		1.26
Di-potassium hydrogen phosphate		0.18
Magnesium chloride (anhydrous)		13.58
Malachite green		0.036

* adjusted as required to meet performance standards

Directions

Suspend 26.75 g in 1 litre of distilled water. Heat gently until dissolved completely. Mix well and distribute into final containers. Sterilize by autoclaving at 115°C for 15 minutes. This medium is very hygroscopic and must be protected from moisture.

Physical Characteristics

Straw/green, free-flowing coarse powder
Colour on reconstitution - blue
Moisture level - less than 7%
pH 5.2 ± 0.2 at 25°C
Clarity - clear

Microbiological Tests Using Optimum Inoculum Dilution

Control Media: Tryptone Soya Agar and XLD Medium

Reactions after incubation at 41 ± 2°C for 24 ± 3 hours

Inoculation with pure cultures

Inoculate 10ml quantities of medium to achieve 1-15 colony-forming units/ml (cfu/ml). Incubate broths at 41 ± 2°C for 24 ± 3 hours. After incubation, subculture onto Tryptone Soya Agar (CM0131) and incubate plates at 37 ± 2°C for 24 ± 3 hours.

Salmonella nottingham	NCTC 7832	1-3mm straw colonies
Salmonella abony	NCTC 6017	1-3mm straw colonies
Salmonella poona	NCTC 4840	1-3mm straw colonies

A satisfactory result is represented by recovery of *Salmonella* strains equal to or greater than a 4 log (10) increase.

Inoculate 10ml quantities of medium to achieve 1E+02 to 1E+04 cfu/ml. Incubate broths at 41 ± 2°C for 24 ± 3 hours. After incubation, subculture onto Tryptone Soya Agar (CM0131) and incubate plates at 37 ± 2°C for 24 ± 3 hours.

Staphylococcus aureus	ATCC® 6538	No growth

Negative strains are inhibited or shall produce at least a 2 log (10) reduction.

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Testing performed in accordance with ISO11133:2014

Reactions after incubation at 41.5 ± 2°C for 24 ± 3 hours

Inoculation with mixed cultures

Inoculate 10ml quantities of medium to achieve 1-10 cfu/ml of *Salmonella* species, to each add 1E+03 to 1E+04 cfu/ml of *Escherichia coli* and *Pseudomonas aeruginosa*. Incubate broths at 41 ± 2°C for 24 ± 3 hours. After incubation, subculture onto XLD Medium (CM0469) and incubate plates at 37 ± 2°C for 24 ± 3 hours.

Salmonella typhimurium	ATCC® 14028	WDCM00031	1-2mm red colonies, black centre
+ *Escherichia coli*	ATCC® 8739	WDCM00012	No growth
+ *Pseudomonas aeruginosa*	ATCC® 27853	WDCM00025	No growth
Salmonella typhimurium	ATCC® 14028	WDCM00031	1-2mm red colonies, black centre
+ *Escherichia coli*	ATCC® 25922	WDCM00013	No growth
+ *Pseudomonas aeruginosa*	ATCC® 27853	WDCM00025	No growth
Salmonella enteritidis	ATCC® 13076	WDCM00030	1-2mm red colonies, black centre
+ *Escherichia coli*	ATCC® 8739	WDCM00012	No growth
+ *Pseudomonas aeruginosa*	ATCC® 27853	WDCM00025	No growth
Salmonella enteritidis	ATCC® 13076	WDCM00030	1-2mm red colonies, black centre
+ *Escherichia coli*	ATCC® 25922	WDCM00013	No growth
+ *Pseudomonas aeruginosa*	ATCC® 27853	WDCM00025	No growth

A satisfactory result is represented by recovery of >100 cfu of *Salmonella* species on XLD Medium (CM0469).

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Inoculation with pure cultures

Inoculate 10ml quantities of medium to achieve 1E+03 to 1E+04 cfu/ml of *Escherichia coli* and *Enterococcus faecalis*. Incubate broths at 41 ± 2°C for 24 ± 3 hours. After incubation, subculture onto Tryptone Soya Agar (CM0131) and incubate plates at 37 ± 2°C for 24 ± 3 hours.

Escherichia coli	ATCC® 8739	WDCM00012	No growth or 1-3mm cream colonies
Escherichia coli	ATCC® 25922	WDCM00013	No growth or 1-3mm cream colonies
Enterococcus faecalis	ATCC® 19433	WDCM00009	No growth or 0.5-1mm straw colonies
Enterococcus faecalis	ATCC® 29212	WDCM00087	No growth or 0.5-1mm straw colonies

A satisfactory result is represented by growth of ≤100 cfu for *Escherichia coli* and <10 cfu for *Enterococcus faecalis* on Tryptone Soya Agar (CM0131).

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Revision History

Section / Step	Description of Change	Reason for Change	Reference
Creation of ISO11133 section	Update to include testing of ISO11133:2014	Change control	BT-CC-1411