

# PVDF Transfer Membrane

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**88520    88518    88585**

Number	Description
88520	PVDF Transfer Membrane, 0.2µm, 26.5cm x 3.75m, 1 roll
88518	PVDF Transfer Membrane, 0.45µm, 26.5cm x 3.75m, 1 roll
88585	PVDF Transfer Membrane, 0.45µm, 10 x 10cm, 10 sheets

**Storage:** Store membranes flat at ambient temperature, away from chemical vapors. Some solvent vapors may partially dissolve the membranes, which will disrupt the pore structure.

## Introduction

Polyvinylidene difluoride (PVDF) membranes are hydrophobic and have high binding affinity for proteins and nucleic acids. These membranes are typically used for applications such as Western, Southern, Northern and dot blots. PVDF membranes offer a better retention of adsorbed proteins than other supports including nitrocellulose.

## Example Procedure for Transferring Proteins to a PVDF Membrane

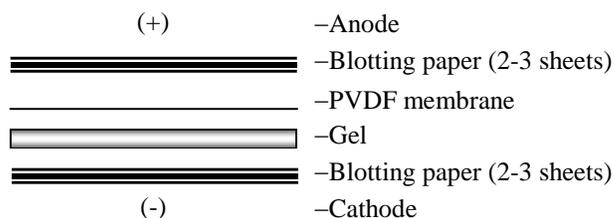
**Note:** Use a sharp instrument to cut the membrane to the size of the gel. Any small tear may result in a larger tear. Always wear gloves when handling PVDF membranes because oils from fingers may prevent proper wetting. Proteins from hands may also bind to the membranes causing background.

### A. Materials required

- Tank transfer system with power supply
- Blotting paper (Part No. 88600)
- Methanol
- Novex™ Tris-Glycine Transfer Buffer (25x) (Part No. LC3675) or other suitable transfer buffer (e.g., 5.8g Tris base, 14.4g glycine, 0.4g SDS (optional), 200mL methanol (100%), pH 8.5. Adjust volume to 1L with ultrapure water).

### B. Method

1. Remove gel from the electrophoresis apparatus and equilibrate in Transfer Buffer for 30 minutes with gentle shaking.  
**Note:** Incubation time is based on a 1.5mm thick gel. Reduce incubation time for thinner gels.
2. Cut membrane to the same dimensions of the gel. Cut a notch in the membrane corner to correspond to a corner of the gel.
3. Wet membrane in 100% methanol for 15 seconds. Ensure that there are no dry areas on the membrane that could inhibit protein transfer.
4. Place membrane in a new container with Transfer Buffer and equilibrate for 15 to 20 minutes.
5. Wet the absorbent filter paper in Transfer Buffer.
6. Use the following component order to form the transfer stack:



7. Connect the leads and perform transfer for 45-90 minutes at 0.8mA/cm<sup>2</sup> of gel.  
**Note:** Transfer time and efficiency will vary depending upon polyacrylamide concentration, gel thickness, the presence of SDS or methanol, pH and ionic strength of the transfer buffer and the molecular weight of the protein. Determine optimal transfer conditions empirically.
8. When the transfer is complete, disconnect leads and disassemble the transfer stack to remove the membrane.
9. Keep membrane moist until ready to use.

## Related Products

<b>XP04200BOX</b>	<b>Novex™ Tris-Glycine protein gels</b> (see <a href="http://thermofisher.com/proteingels">thermofisher.com/proteingels</a> for a complete listing)
<b>NW04120BOX</b>	<b>Bolt™ Bis-Tris Plus protein gels</b> (see <a href="http://thermofisher.com/proteingels">thermofisher.com/proteingels</a> for a complete listing)
<b>LC5615</b>	<b>iBright™ Prestained Protein Ladder</b>
<b>24585</b>	<b>Pierce™ Reversible Protein Stain Kit for PVDF Membranes</b> , sufficient material for 10 mini blots
<b>88600</b>	<b>Western Blotting Filter Paper, 8cm × 10.5cm</b> , 100 sheets
<b>34580</b>	<b>SuperSignal™ West Pico Plus Chemiluminescent Substrate</b> , 500mL
<b>34075</b>	<b>SuperSignal West Dura Extended Duration Substrate</b> , 100mL
<b>34095</b>	<b>SuperSignal West Femto Maximum Sensitivity Substrate</b> , 100mL
<b>21059</b>	<b>Restore™ Western Blot Stripping Buffer</b> , 500mL
<b>34090</b>	<b>CL-XPosure™ Film (5" × 7")</b> , 100 sheets
<b>34091</b>	<b>CL-XPosure Film (8" × 10")</b> , 100 sheets
<b>21065</b>	<b>Pierce Background Eliminator Kit</b> , for eliminating background from X-ray film

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