

Denaturing Formaldehyde Gels in MOPS Buffer

This protocol is for the Denaturing Formaldehyde Gels in MOPS Buffer

1. Freshly prepare 10X MOPS buffer: 0.4 M MOPS (pH 7.0), 0.1 M sodium acetate, 0.01 M EDTA (pH 8.0).
2. Prepare 1% TopVision™ Agarose gel as follows:
 - stir 1g of agarose powder in 72 ml of deionized water;
 - melt the agarose, and then add 10 ml of 10X MOPS buffer and mix;
 - when the agarose solution cools to 60°C, add 18 ml of fresh formaldehyde (37%) in a fume hood and mix thoroughly;
 - pour the gel.
3. Place the gel into an electrophoresis apparatus containing 1X MOPS buffer.
4. Heat the RNA samples and ladder at 70°C for 10 min, and then chill on ice for 3 min.
5. Load onto the gel.

Note

There is no need to stain the gel as ethidium bromide present in 2X RNA Loading Dye is sufficient for visualization under UV light.

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