



# Stimulation and expansion of human B cells by magnetic bead-based activation of CD4<sup>+</sup> T cells

## Highlights

- Use of Dynabeads Human T-Activator CD3/CD28 beads to stimulate B cells and differentiate them into antibody-producing plasma cells or memory B cells via autologous stimulation of CD4<sup>+</sup> Th cells
- A Dynabeads magnetic bead-based activation method for the study of neoplasms (multiple myeloma and chronic lymphocytic leukemia) and antibody research to expand and proliferate B and plasma cells

## Keywords

B cells, Th cells, Dynabeads Untouched Human B Cells, Dynabeads Untouched Human CD4, Dynabeads Human T-Activator CD3/CD28, plasma cells, memory B cells



## Introduction

B cells are part of the acquired immune response to diseases. These lymphocytes produce antibodies against a wide range of pathogens and orchestrate pro-inflammatory responses. Only B cells that differentiate into large plasma cells will produce antibodies. In addition, some B cells will differentiate into memory B cells.

The dependency on Th cell activation is not only confined to normal B cell differentiation; it also has been reported in proliferation of certain blood cancers, such as multiple myeloma [1] and chronic lymphocytic leukemia [2]. These reports and findings show Th cell–B cell interaction and crosstalk are not only critical for B cell development but also necessary for the progression of some blood cancers.

Th cell activation and differentiation of B cells to plasma cells is crucial for the collective understanding of many cancers of the blood. Therefore, a method for expansion of B cells *in vitro* is the cornerstone to study mechanisms of drug resistance, proliferation pathways, oncogenes, and neoantigens. Here we describe a method to isolate Th cells and B cells using negative isolation (untouched) to leave important biomarkers free for downstream applications. The use of Gibco™ Dynabeads™ Human T-Activator CD3/CD28 beads is crucial for stimulating Th cells into driving B cell differentiation and proliferation.

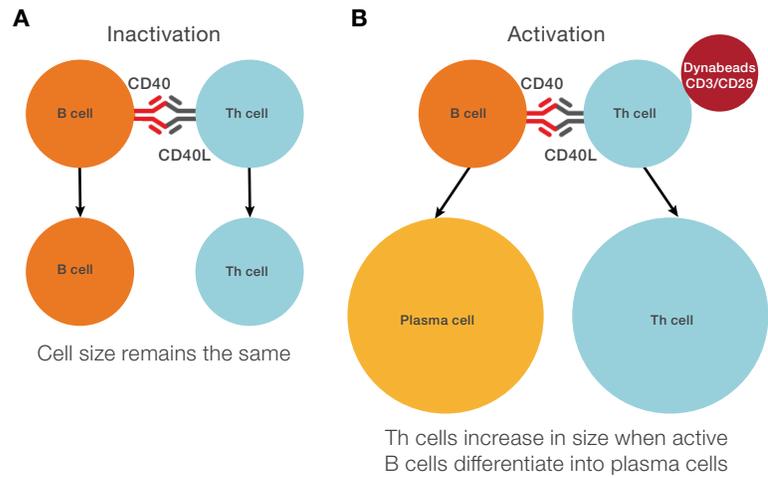
**Activation of Th cells using Dynabeads CD3/CD28 beads is enough to stimulate proliferation of B cells *in vitro***

To measure cell proliferation, the nuclear protein Ki-67 was used as a biomarker. In addition, forward scatter (FSC) area was used to address cell size for B cells differentiating into plasma cells. Figure 1 illustrates the interaction between a naïve B cell in resting state with an activated Th cell. Th cells are activated *in situ* through the co-stimulatory domains CD3 and CD28 by an antigen presenting cell. In turn, this activates naïve B cells into antibody-secreting plasma cells or memory B cells.

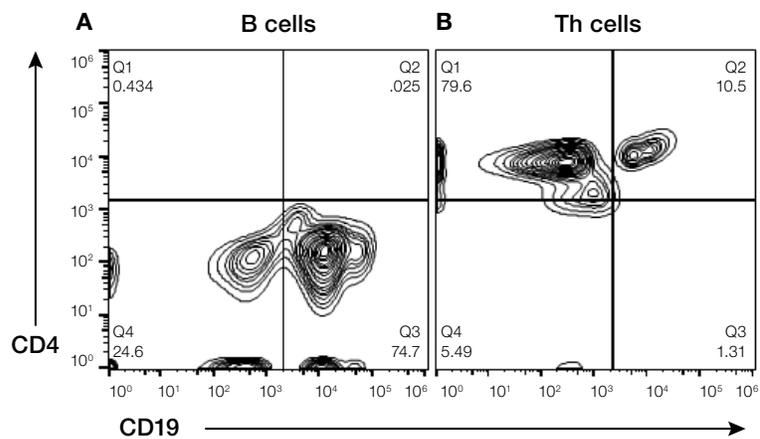
Th experimental design was set up to co-culture a ratio of 1:1 B cells to Th cells. Test cells were cultured with either 200 U/mL IL-2 alone or Dynabeads Human T-Activator CD3/CD28 beads alone, at a ratio of 1:1 beads:cells. Positive controls were incubated with a combination of 200 U/mL IL-2 and Dynabeads Human T-Activator CD3/CD28 beads at a ratio of 1:1 beads:cells. Negative controls were B cells alone without stimulants, Th cells alone without stimulants, and a co-culture of 1:1 B cells:Th cells without stimulants.

CD4<sup>+</sup> Th cells and CD19<sup>+</sup> B cells were negatively selected, leaving the cells' surfaces untouched by antibodies and beads. This resulted in a purity of ~75% CD19<sup>+</sup> B cells and ~80% CD4<sup>+</sup> Th cells (Figure 2).

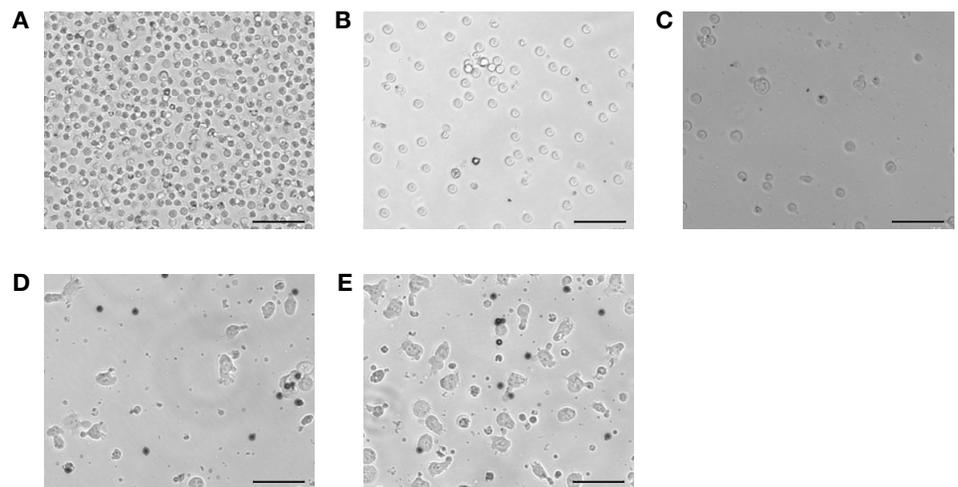
B cells differentiated into plasma cells after Th cells were stimulated using Dynabeads Human T-Activator CD3/CD28 beads (Figures 3 and 4). Light microscopy images in Figure 3 show the differences in cell sizes between resting naïve B cells (Figure 3A) and blasting plasma cells (rounded cells; Figures 3D and 3E). Figure 4 shows a blasting B cell (Figures 4C and 4G) that is proliferating following Th cell stimulation (Figures 4D and 4H).



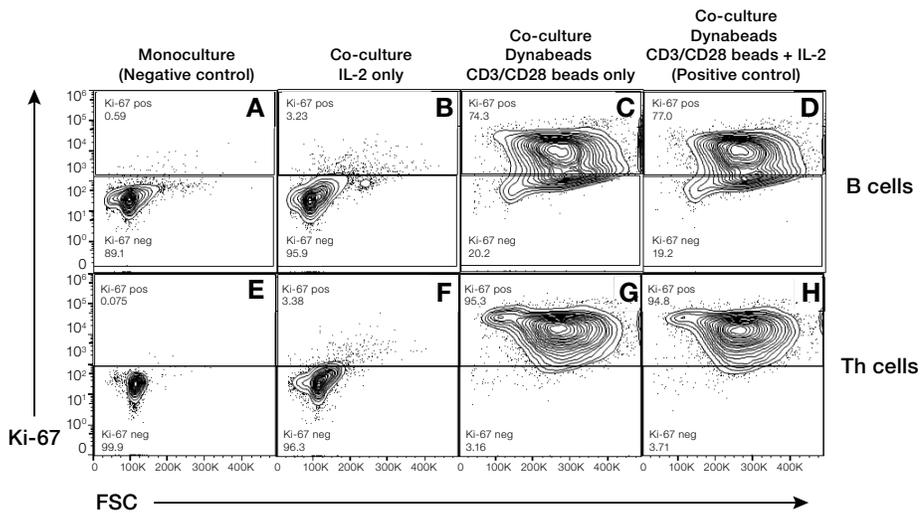
**Figure 1. Activated Th cells expand and proliferate B cells to plasma cells. (A)** Inactivated Th cells are unable to stimulate naïve B cells into a proliferative state morphology. **(B)** Upon activation of Th cells by Dynabeads Human T-Activator CD3/CD28 beads, Th cells were able to drive naïve B cells into plasma cells.



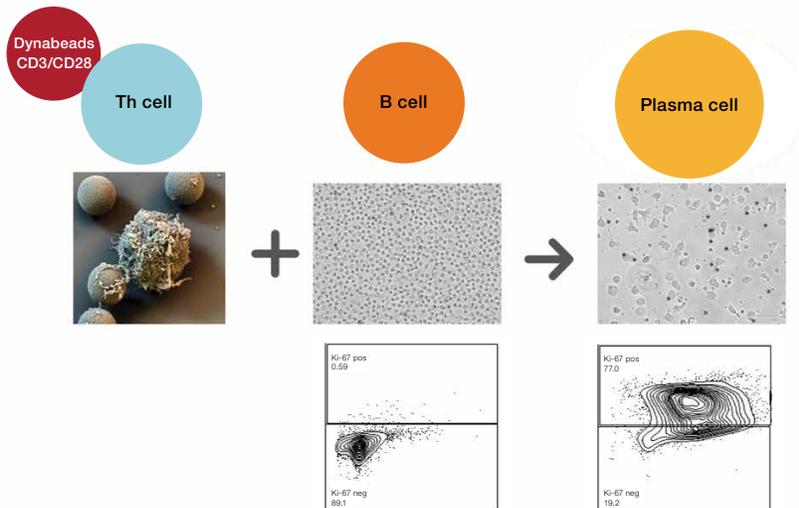
**Figure 2. Isolation of CD19<sup>+</sup> B cells and CD4<sup>+</sup> Th cells using the Invitrogen™ Dynabeads™ Untouched Human B Cells Kit and Dynabeads Untouched Human CD4 T Cells Kit. (A)** Gated B cells following negative isolation were 74.7% of the total live cell population positive for the biomarker CD19 (Q3). **(B)** Th cells gated on total live cells were 79.6% pure following negative isolation (Q1).



**Figure 3. Light images of cells after four days in culture. (A)** B cells without stimulants (negative control). **(B)** Th cells without stimulants (negative control). **(C)** Co-culture of both B and Th cells with 200 U/mL IL-2. **(D)** Co-culture of both B and Th cells with Dynabeads Human T-Activator CD3/CD28 beads. **(E)** Co-culture of B and Th cells treated with 200 U/mL IL-2 and Dynabeads Human T-Activator CD3/CD28 beads. Scale bar is 50 μm.



**Figure 4. Cell proliferation measured by FSC and Ki-67.** Cultures of B cells or Th cells (A and E) were left unstimulated, and they remained small with no Ki-67 expression, as expected. For cells in co-culture with IL-2 stimulation (B and F), minimal Ki-67 expression and blasting of B cells were observed. For cells in co-culture with Dynabeads CD3/CD28 beads (C and G) and cells in co-culture with Dynabeads CD3/CD28 beads and IL-2 (D and H), B cells were able to blast into larger cells after activation of Th cells. These B cells had high Ki-67 expression and increased cell sizes.



**Figure 5. Summary of B cell activation and stimulation to a plasma cell using Dynabeads Human T-Activator CD3/CD28 beads.** By activation of Th cells with Dynabeads Human T-Activator CD3/CD28 beads, naïve B cells were able to blast into larger memory B cells or plasma cells.

## Summary

The results shown here demonstrate the importance of Th cells stimulation and activation of B cells *in vitro* (Figure 5). In addition, we show that using Dynabeads Human T-Activator CD3/CD28 beads alone were able to stimulate the Th cells into proliferating and activating B cells without IL-2.

This method can be adopted to expand and test cancerous B cells, such as multiple myeloma cells. It can also be used to blast naïve B cells into antibody-producing plasma cells for experimental CAR B cell research. And finally, as CD19 is becoming an important target in cancerous B cells, using the Dynabeads Untouched Human B Cells Kit prohibits CD19 on B cells to be blocked with beads or antibodies.

## Authors

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## References

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## Ordering information

Product	Cat. No.
Dynabeads Untouched Human B Cells Kit	<a href="#">11351D</a>
Dynabeads Untouched Human CD4 T Cells Kit	<a href="#">11346D</a>
Dynabeads Human T-Activator CD3/CD28	<a href="#">11131D</a>
eBioscience CD4 Monoclonal Antibody, PE	<a href="#">12-0047-42</a>
eBioscience CD19 Monoclonal Antibody, APC	<a href="#">17-0199-42</a>
Ki-67 Monoclonal Antibody, FITC	<a href="#">MHK16701</a>
SYTOX Blue Dead Cell Stain	<a href="#">S34857</a>

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