

PRODUCT INFORMATION

DNase I, RNase-free

#EN0521 1000 u

Lot: Expiry Date:

Concentration: 1 u/μl

Supplied with: 1 ml of 10X Reaction Buffer with MgCl₂
1 ml of 50 mM EDTA

Store at -20°C

www.thermoscientific.com/fermentas**Description**

DNase I is an endonuclease that digests single- and double-stranded DNA. It hydrolyzes phosphodiester bonds producing mono- and oligodeoxyribonucleotides with 5'-phosphate and 3'-OH groups.

The enzyme activity is strictly dependent on Ca²⁺ and is activated by Mg²⁺ or Mn²⁺ ions:

- in the presence of Mg²⁺, DNase I cleaves each strand of dsDNA independently, in a statistically random fashion (1);
- in the presence of Mn²⁺, the enzyme cleaves both DNA strands at approximately the same site, producing DNA fragments with blunt ends or with one or two nucleotide overhangs (1).

Applications

- Preparation of DNA-free RNA (1).
- Removal of template DNA following *in vitro* transcription (1), *see* protocol on reverse page.
- Preparation of DNA-free RNA prior to RT-PCR and RT-qPCR (2), *see* protocol on reverse page.
- DNA labeling by nick-translation in conjunction with DNA Polymerase I (1), *see* protocol on reverse page.
- Studies of DNA-protein interactions by DNase I, RNase-free footprinting (1).
- Generation of a library of randomly overlapping DNA inserts. Reaction buffer containing Mn²⁺ is used (3).

Rev.9



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Source

E. coli cells with a cloned gene encoding bovine DNase I.

Molecular Weight

29 kDa monomer.

Definition of Activity Unit

One unit of the enzyme completely degrades 1 μg of plasmid DNA in 10 min at 37°C.

Enzyme activity is assayed in the following mixture:

10 mM Tris-HCl (pH 7.5 at 25°C), 2.5 mM MgCl₂, 0.1 mM CaCl₂, 1 μg of pUC19 DNA. One DNase I unit is equivalent to 0.3 Kunitz unit (4).

Storage Buffer

The enzyme is supplied in: 50 mM Tris-HCl (pH 7.5), 10 mM CaCl₂ and 50% (v/v) glycerol.

10X Reaction Buffer with MgCl₂

100 mM Tris-HCl (pH 7.5 at 25°C), 25 mM MgCl₂, 1 mM CaCl₂.

Inhibition and Inactivation

- Inhibitors: metal chelators, transition metals (e.g., Zn) in millimolar concentrations, SDS (even at concentrations less than 0.1%), reducing agents (DTT and β-mercaptoethanol), ionic strength above 50-100 mM.
- Inactivated by heating at 65°C for 10 min in the presence of EGTA or EDTA (use at least 1 mol of EGTA/EDTA per 1 mol of Mn²⁺/Mg²⁺ (5)).

Note

DNase I is sensitive to physical denaturation. Mix gently by inverting the tube. Do not vortex.

CERTIFICATE OF ANALYSIS**Ribonuclease Assay**

No degradation of RNA was observed after incubation of 5 units of DNase I with 160 ng RNA for 4 hours at 37°C.

Quality authorized by:

Jurgita Zilinskiene

(continued on reverse page)

Removal of genomic DNA from RNA preparations

1. Add to an RNase-free tube:

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|--|----------|
| RNA | 1 µg |
| 10X reaction buffer with MgCl ₂ | 1 µl |
| DNase I, RNase-free | 1 u |
| DEPC-treated Water (#R0601) | to 10 µl |

2. Incubate at 37°C for 30 min.

3. Add 1 µl 50 mM EDTA and incubate at 65°C for 10 min. RNA hydrolyzes during heating with divalent cations in the absence of a chelating agent (5). Alternatively, use phenol/chloroform extraction.

4. Use the prepared RNA as a template for reverse transcriptase.

Note

- Do not use more than 1 u of DNase I, RNase-free per 1 µg of RNA.
- If using DNase I, HC, enzyme can be diluted in 1X DNase reaction buffer just prior to use, or in storage buffer (not supplied *see composition on reverse page*) for longer storage.
- Volumes of the reaction mixture and 50 mM EDTA solution can be scaled up for larger amounts of RNA. The recommended final concentration of RNA is 0.1 µg/µl.
- Thermo Scientific RiboLock RNase Inhibitor (#E00381), typically at 1 u/µl, can also be included in the reaction mixture to prevent RNA degradation.

Removal of template DNA after *in vitro* transcription

1. Add 2 u of DNase I, RNase-free per 1 µg of template DNA directly to a transcription reaction mixture. In some cases, the amount of enzyme should be determined empirically.

2. Incubate at 37°C for 15 minutes.

3. Inactivate DNase I by phenol/chloroform extraction.

DNA labeling by nick-translation

1. Mix the following components:

| | |
|---|---------------------------|
| 10X reaction buffer for DNA Polymerase I | 2.5 µl |
| Mixture of 3 dNTPs, 1 mM* each (without the labeled dNTP) | 1.25 µl |
| [α- ³² P]-dNTP, ~110 TBq/mmol (3000 Ci/mmol) | 1.85-3.7 MBq (50-100 µCi) |
| DNase I, RNase-free freshly diluted to 0.002 u/µl** | 1 µl |
| DNA Polymerase I (#EP0041) | 0.5-1.5 µl (5-15 u) |
| Template DNA | 0.25 µg |
| Water, nuclease-free (#R0581) | to 25 µl |

2. Immediately incubate at 15°C for 15-60 min.

3. Terminate the reaction by adding 1 µl of 0.5 M EDTA, pH 8.0 (#R1021).

4. Take an aliquot (1 µl) to determine the efficiency of label incorporation. A specific activity of at least 10⁸ cpm/µg DNA is expected.

Note

- The labeled DNA can be purified from the unincorporated labelled dNTPs using Thermo Scientific GeneJET PCR Purification Kit (#K0701).
- * To prepare a mixture of 3 non-labeled dNTPs (1 mM of each), mix 1 µl aliquots of stock solutions of each dNTP (100 mM, from #R0181) with 97 µl of Water, nuclease-free (#R0581). Store at -20°C.
- ** DNase I, RNase-free can be diluted with 1X reaction buffer for DNA Polymerase I: 50 mM Tris-HCl (pH 7.5 at 25°C), 10 mM MgCl₂ and 1 mM DTT.

References

1. Sambrook, J., Russell, D.W., Molecular Cloning: A Laboratory Manual, the third edition, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York, 2001.
2. Kienzle, N., et al., DNase I treatment is a prerequisite for the amplification of cDNA from episomal-based genes, *BioTechniques*, 20, 612-616, 1996.
3. Anderson, S., Shotgun DNA sequencing using cloned DNase I-generated fragments, *Nucleic Acids Res.*, 9, 3015-3027, 1981.
4. Kunitz, M., *J.Gen.Physiol.*, 33, 349-362, 1950.
5. Wiame, I., et al., Irreversible heat inactivation of DNase I without RNA degradation, *BioTechniques*, 29, 252-256, 2000.

PRODUCT USE LIMITATION

This product is developed, designed and sold exclusively *for research purposes and in vitro use only*. The product was not tested for use in diagnostics or for drug development, nor is it suitable for administration to humans or animals. Please refer to www.thermoscientific.com/fermentas for Material Safety Data Sheet of the product.