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### OXOID QUALITY ASSURANCE PRODUCT SPECIFICATION

### PERFRINGENS AGAR BASE (TSC & SFP) CM0587

PERFRINGENS AGAR BASE (TSC & SFP)		CM0587	
Typical Formula*			
Tryptose	grams per litre	15.0	
Soya peptone		5.0	
Yeast extract		5.0	
Sodium metabisulphite		1.0	
Ferric ammonium citrate		1.0	
Agar		19.0	

<sup>\*</sup> adjusted as required to meet performance standards

#### **Directions**

Suspend 23 grams in 500ml of distilled water and heat gently until the agar is completely dissolved. Sterilize by autoclaving at 121°C for 10 minutes. Allow the medium to cool to 50°C and add 25ml of Egg Yolk Emulsion (SR0047) and the rehydrated contents of one vial of TSC Supplement (SR0088) or SFP Supplement (SR0093). Mix and pour plates.

#### **Physical Characteristics**

Straw, free-flowing powder
Colour on reconstitution - straw/green
Moisture level - less than or equal to 7%
pH - 7.6 ± 0.2 at 25°C
Clarity - clear
Gel strength - firm, comparable to 19g/litre of agar

### **Microbiological Tests Using Optimum Inoculum Dilution**

Control Media: Tryptone Soya Agar or Columbia Blood Agar Base enriched with 5% v/v horse blood, where appropriate



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### **OXOID QUALITY ASSURANCE PRODUCT SPECIFICATION**

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#### Reactions after incubation at 37 ± 2°C for 24 hours under anaerobic conditions

Tested with the addition of TSC Supplement SR0088 and Egg Yolk Emulsion SR0047

#### Pour plate technique

Medium is challenged with 10-100 colony-forming units

Clostridium perfringens ATCC®13124 1-3mm black colonies with halo Clostridium bifermentans NCTC506 0.5-2mm black colonies with halo

Plates overlaid with TSC Agar without egg yolk.

A satisfactory result is represented by recovery equal to or greater than 70% of the control medium.

For *Clostridium bifermentans* NCTC506, a satisfactory result is represented by recovery equal to or greater than 30% of the control medium.

#### Surface plate technique

Medium is challenged with 1E+04 to 1E+06 colony-forming units

Clostridium tetani ATCC®9441 No growth

Proteus mirabilis ATCC®29906 No growth or ppt-3mm translucent colonies

Negative strains are inhibited or shall produce at least a 4 log (10) reduction.

### Testing has been performed in accordance with ISO11133:2014

### <u>Table E</u>

ISO Standard 7937 tested with the addition of TSC Supplement SR0088

### Reactions after incubation at 37 ± 2°C for 20 ± 2 hours under anaerobic conditions

### Pour plate technique

Medium is challenged with 50-120 colony-forming units

Clostridium perfringens	ATCC®13124	WDCM00007	1-3mm black colonies
Clostridium perfringens	ATCC®12916	WDCM00080	1-3mm black colonies



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Plates overlaid with TSC Agar.

A satisfactory result is represented by recovery equal to or greater than 50% of the control medium.

### Surface plate technique

Medium is challenged with 1E+04 to 1E+06 colony-forming units

Escherichia coli ATCC®25922 WDCM00013 No growth Escherichia coli ATCC®8739 WDCM00012 No growth

Negative strains are inhibited.

ISO Standard 15213 tested using unsupplemented medium

### Reactions after incubation at 37 ± 2°C for 48 ± 2 hours under anaerobic conditions

Medium is challenged with 50-120 colony-forming units

### Pour plate technique

Clostridium perfringens	ATCC®13124	WDCM00007	1-3mm black colonies
Clostridium perfringens	ATCC®12916	WDCM00080	1-3mm black colonies

A satisfactory result is represented by recovery equal to or greater than 70% of the control medium.

### Surface plate technique

Medium is challenged with 1E+04 to 1E+06 colony-forming units

Escherichia coli	ATCC®25922	WDCM00013	0.25-1mm white/cream colonies
Escherichia coli	ATCC®8739	WDCM00012	0.25-1mm white/cream colonies

### Table F

ISO Standard 14189 tested with the addition of TSC Supplement SR0088

#### Reactions after incubation at 44 ± 2°C for 21 ± 3 hours under anaerobic conditions

#### Filtration technique

Medium is challenged with 50-120 colony-forming units

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Clostridium perfringensATCC®13124WDCM000071-3mm black coloniesClostridium perfringensATCC®12916WDCM000801-3mm black coloniesClostridium perfringensATCC®10543WDCM001741-3mm black colonies

Plates overlaid with TSC Agar.

A satisfactory result is represented by recovery equal to or greater than 50% of the control medium.

Medium is challenged with 1E+04 to 1E+05 colony-forming units

Bacillus subtilis ATCC®6633 WDCM00003 No growth

Negative strains are inhibited

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## **OXOID QUALITY ASSURANCE PRODUCT SPECIFICATION**

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# **Revision History**

Section / Step	Description of Change	Reason for Change	Reference
Creation of ISO11133 section	Update to include testing of ISO11133:2014	Change control	BT-CC-1358
Tested with the addition of TSC Supplement SR0088 and Egg Yolk Emulsion SR0047	Change of <i>Proteus</i> spp. from complete inhibition to 4 log (10) reduction	Change control	BT-CC-2226