

	Document Owner Department: QC	MBD-BT-SPEC-0455
		Page 1 of 3
<b>OXOID QUALITY ASSURANCE PRODUCT SPECIFICATION</b>		
<b>PERFRINGENS (TSC) SELECTIVE SUPPLEMENT SR0088E</b>		

**PERFRINGENS (TSC) SELECTIVE SUPPLEMENT**

**SR0088E**

**Formula**

Per vial (each vial is sufficient to supplement 500ml of medium)

D-cycloserine 200.0 mg

**Description**

A selective supplement for the isolation of *Clostridium perfringens* when used with Perfringens Agar Base (TSC/SFP) CM0587.

**Directions**

To one vial add 2ml of sterile distilled water and dissolve the contents completely. Avoid frothing of solution. Add the contents aseptically to 500ml Perfringens Agar Base (TSC/SFP) CM0587 cooled to 50°C together with 25ml of Egg Yolk Emulsion (SR0047). Mix well and pour into sterile Petri dishes.

**Physical Characteristics**

White pellet  
 Colour after reconstitution – colourless or pale yellow  
 Sterility - passes test

**Microbiological Tests Using Optimum Inoculum Dilution**

Control Media: Tryptone Soya Agar or Columbia Blood Agar Base enriched with 5% v/v horse blood, where appropriate

**Reactions after incubation at 37 ± 2°C for 24 hours under anaerobic conditions**

Tested in Perfringens Agar Base (TSC/SFP) CM0587 with the addition of 5% v/v Egg Yolk Emulsion (SR0047)

**Pour plate technique**

Medium is challenged with 10-100 colony-forming units

<i>Clostridium perfringens</i>	ATCC®13124	1-3mm black colonies with halo
<i>Clostridium bifermentans</i>	NCTC506	0.5-2mm black colonies with halo

Plates overlaid with TSC Agar without egg yolk.

	Document Owner Department: QC	MBD-BT-SPEC-0455
		Page 2 of 3
<b>OXOID QUALITY ASSURANCE PRODUCT SPECIFICATION</b>		
<b>PERFRINGENS (TSC) SELECTIVE SUPPLEMENT SR0088E</b>		

A satisfactory result is represented by recovery equal to or greater than 70% of the control medium.

For *Clostridium bifermentans* NCTC506, a satisfactory result is represented by recovery equal to or greater than 30% of the control medium.

### Surface plate technique

Medium is challenged with 1E+04 to 1E+06 colony forming units

<i>Clostridium tetani</i>	ATCC®9441	No growth
<i>Proteus mirabilis</i>	ATCC®29906	No growth or ppt-3mm translucent colonies

Negative strains are inhibited or shall produce at least a 4 log (10) reduction.

### Testing has been performed in accordance with ISO11133:2014

#### Table E

ISO Standard 7937 tested in Perfringens Agar Base (TSC/SFP) CM0587

### Reactions after incubation at 37 ± 2°C for 20 ± 2 hours under anaerobic conditions

#### Pour plate technique

Medium is challenged with 50-120 colony-forming units

<i>Clostridium perfringens</i>	ATCC®13124	WDCM00007	1-3mm black colonies
<i>Clostridium perfringens</i>	ATCC®12916	WDCM00080	1-3mm black colonies

Plates overlaid with TSC Agar.

A satisfactory result is represented by recovery equal to or greater than 50% of the control medium.

### Surface plate technique

Medium is challenged with 1E+04 to 1E+06 colony-forming units

<i>Escherichia coli</i>	ATCC®25922	WDCM00013	No growth
<i>Escherichia coli</i>	ATCC®8739	WDCM00012	No growth

	Document Owner Department: QC	MBD-BT-SPEC-0455
		Page 3 of 3
<b>OXOID QUALITY ASSURANCE PRODUCT SPECIFICATION</b>		
<b>PERFRINGENS (TSC) SELECTIVE SUPPLEMENT SR0088E</b>		

Negative strains are inhibited

Table F

ISO Standard 14189 tested in Perfringens Agar Base (TSC/SFP) CM0587

**Reactions after incubation at 44 ± 2°C for 21 ± 3 hours under anaerobic conditions**

**Filtration technique**

Medium is challenged with 50-120 colony-forming units

<i>Clostridium perfringens</i>	ATCC®13124	WDCM00007	1-3mm black colonies
<i>Clostridium perfringens</i>	ATCC®12916	WDCM00080	1-3mm black colonies
<i>Clostridium perfringens</i>	ATCC®10543	WDCM00174	1-3mm black colonies

Plates overlaid with TSC Agar.

A satisfactory result is represented by recovery equal to or greater than 50% of the control medium.

Medium is challenged with 1E+04 to 1E+05 colony-forming units

<i>Bacillus subtilis</i>	ATCC®6633	WDCM00003	No growth
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Negative strains are inhibited

**Revision History**

Section / Step	Description of Change	Reason for Change	Reference
Physical characteristics	Added colour of solution on reconstitution	Change control	MOC-2023-0303