TaqMan[™] RNA-to-C_T[™] 1-Step Kit

Catalog Numbers 4392653, 4392938, and 4392656

Pub. No. 4392668 Rev. C

Note: For safety and biohazard guidelines, see the "Safety" appendix in the following product documentation: *TaqMan* RNA-to-C_T RNA-to-C_T appendix in the following product documentation: *TaqMan* RNA-to-C_T appropriate protective eyewear, clothing, and gloves.

Product description

Use the TaqMan[™] RNA-to- $C_T^{\text{\tiny TM}}$ 1-Step Kit to perform one step RT-PCR with TaqMan[™] reagents for quantification experiments on a real-time PCR system.

Contents and storage

Contents	Cat. No. 4392653 (40 × 50 µL reactions)	Cat. No. 4392938 (200 × 50 μL reactions)	Cat. No. 4392656 (2,000 × 50 µL reactions)	Storage ^[1]
2X TaqMan™ RT-PCR Mix	1 mL	5 mL	10 × 5 mL	-25°C to -15°C on receipt, protect from light 2-8°C after first use, protect from light
40X TaqMan™ RT Enzyme Mix	50 μL	250 μL	10 × 250 μL	–25°C to −15°C

^[1] See packaging for expiration date.

Methods

Before you begin

- Thoroughly mix the 2X TaqMan[™] RT-PCR Mix. Do not create excess bubbles.
- Thoroughly mix the 40X TagMan™ RT Enzyme Mix, then briefly centrifuge to resuspend. Do not create excess bubbles.
- Determine the total number of RT-PCR reactions required. We recommend performing four replicates of each reaction.

Prepare the RT-PCR Reaction Mix

1. Combine the following components for the number of reactions required, plus 10% overage.

Component	Volume per reaction			
Component	384-well plate	96-well (0.1-mL) plate	96-well (0.2-mL) plate	
2X TaqMan™ RT-PCR Mix	5 µL	10 μL	25 μL	
40X TaqMan™ RT Enzyme Mix	0.25 μL	0.5 µL	1.25 µL	
20X TaqMan™ Gene Expression Assay	0.5 μL	1 µL	2.5 μL	
RNA template	Variable	Variable	Variable	
Nuclease-free water	Variable	Variable	Variable	
Total RT-PCR Reaction Mix volume per reaction	10 μL	20 μL	50 μL	

- 2. Vortex briefly to mix.
- 3. Centrifuge the tubes briefly to spin down the contents and eliminate any air bubbles.



Prepare the RT-PCR reaction plate

1. Transfer the appropriate volume of RT-PCR Reaction Mix to each well of the plate.

384-well plate: 10 µL
96-well 0.1-mL plate: 20 µL
96-well 0.2-mL plate: 50 µL

2. Seal the reaction plate, then centrifuge briefly to bring the RT-PCR Reaction Mix to the bottom of the wells and eliminate air bubbles.

Run the RT-PCR reactions

See the appropriate instrument user guide for detailed instructions to program the thermal-cycling conditions or to run the plate.

1. Set up a plate document or experiment file using the following conditions:

Instrument	Step	Temperature	Time	Cycles
StepOne™ Real-Time PCR System	Reverse transcription	48°C	15 minutes	1
StepOnePlus™ Real-Time PCR System	Enzyme activation	95°C	10 minutes	1
QuantStudio™ 3 or 5 Real-Time PCR System	Denaturation	95°C	15 seconds	
QuantStudio™ 6 or 7 Flex Real-Time PCR System				
QuantStudio™ 6 Pro or 7 Pro Real-Time PCR System				
QuantStudio™ 12K Flex Real–Time PCR System	Annealing/extension	60°C	1 minute	40
7500 Real-Time PCR System				
7500 Fast Real-Time PCR System				
7900HT Real-Time PCR System				

2. Select Standard cycling mode.

IMPORTANT! TaqMan[™] RNA-to-C_T[™] 1-Step Kit does not support the fast cycling mode. Use standard cycling mode to run the RT-PCR reactions.

- 3. Enter the sample volume.
- 4. Load the reaction plate.
- 5. Start the run.

Guidelines for data analysis

Data analysis varies depending on the instrument used. Refer to the $TaqMan^{\top}RNA$ -to- $C_T^{\top}1$ -Step Kit User Guide (Pub. No. 4393463) and your instrument documentation for detailed information on data analysis.

Limited product warranty

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For descriptions of symbols on product labels or product documents, go to thermofisher.com/symbols-definition.

Revision history: Pub. No. 4392668

Revision	Date	Description	
	 The storage conditions for the 2X TaqMan[™] RT-PCR Mix and the 40X TaqMan[™] RT Enzyme Mix were updated. 		
С	C 22 February 2023	The volumes per reaction for the RT-PCR Reaction Mix were updated.	
		The real-time PCR instrument list was updated.	
В	15 October 2018	Updated for manufacturer, general style, formatting, and branding.	
А	26 September 2007	New document.	

The information in this guide is subject to change without notice.

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