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OXOID QUALITY ASSURANCE PRODUCT SPECIFICATION		
VIOLET RED BILE GLUCOSE AGAR (ISO) CM1082		

VIOLET RED BILE GLUCOSE AGAR (ISO)

CM1082

Typical Formula*

	grams per litre	
Yeast extract		3.0
Enzymatic digest of animal tissues		7.0
Sodium chloride		5.0
Bile salts No.3		1.5
Glucose		10.0
Neutral red		0.03
Crystal violet		0.002
Agar		12.0

* adjusted as required to meet performance standards

Directions

Suspend 38.5g in 1 litre of distilled water. With frequent agitation, bring to the boil to dissolve completely. Cool to 50°C. Mix well and pour into sterile Petri dishes or hold at 45°C when using the pour plate technique. DO NOT AUTOCLAVE.

Physical Characteristics

Straw/pink, free-flowing powder
 Colour on reconstitution - purple
 Moisture level - less than or equal to 7%
 pH 7.4 ± 0.2 at 25°C
 Clarity - clear
 Gel strength - firm, comparable to 12.0g/litre of agar

Microbiological Tests Using Optimum Inoculum Dilution

Control Medium: Tryptone Soya Agar

Reactions after incubation at 37 ± 2°C for 24 ± 2 hours

Inoculation using pour plate technique

Medium is challenged with 50-150 colony-forming units

<i>Klebsiella pneumoniae</i>	ATCC®29665	1-2mm purple/pink colonies and halo
<i>Proteus mirabilis</i>	ATCC®12453	0.5-2mm purple colonies with/without halo
<i>Enterobacter aerogenes</i>	ATCC®13048	1-4mm purple/pink colonies with/without halo
<i>Shigella sonnei</i>	ATCC®25931	1-3mm irregular, purple/pink colonies with/without halo

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A satisfactory result for pour plate technique is represented by recovery of positive strains equal to or greater than 50% of the control medium.

There shall be no gassing in the medium.

Inoculation using surface plate technique

Medium is challenged with 50-150 colony-forming units

Pseudomonas aeruginosa ATCC®9027 1-3mm straw colonies, no halo

For *Pseudomonas aeruginosa* ATCC®9027, a satisfactory result for surface plate technique is represented by recovery equal to or greater than 50% of the control medium.

Medium is challenged with 1E+04 to 1E+06 colony-forming units

Proteus mirabilis ATCC®12453 0.5-2mm purple/pink colonies, no swarming

Testing performed in accordance with ISO11133: 2014

Reactions after incubation at 37 ± 2°C for 24 ± 2 hours

Inoculation using pour plate technique

Medium is challenged with 50-120 colony-forming units

<i>Escherichia coli</i>	ATCC®8739	WDCM00012	1-3mm purple/pink colonies and halo
<i>Escherichia coli</i>	ATCC®25922	WDCM00013	1-3mm purple/pink colonies with/without halo
<i>Salmonella typhimurium</i>	ATCC®14028	WDCM00031	0.5-2mm purple/pink colonies with/without halo
<i>Salmonella enteritidis</i>	ATCC®13076	WDCM00030	0.5-2mm purple/pink colonies with/without halo

A satisfactory result for pour plate technique is represented by recovery of positive strains equal to or greater than 50% of the control medium.

There shall be no gassing in the medium.

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Selectivity determined by qualitative testing based on the methods described in ISO 11133:2014

Reactions after incubation at 37 ± 2°C for 24 ± 2 hours

Inoculation using surface plate technique

Medium is challenged with 1E+04 to 1E+06 colony-forming units

Enterococcus faecalis ATCC®19433 WDCM00009 No growth to pinpoint colonies

Enterococcus faecalis ATCC®29212 WDCM00087 No growth to pinpoint colonies

Negative strains are inhibited or produce a negative diagnostic reaction.

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Revision History

Section / Step	Description of Change	Reason for Change	Reference
Entire document	Update to new template. Correction of typographical/minor errors.	N/A	N/A
Microbiological characteristics	<i>Enterococcus faecalis</i> growth	Change control	MOC-2023-0574