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OXOID QUALITY ASSURANCE PRODUCT SPECIFICATION		
VIOLET RED BILE LACTOSE AGAR CM0107		

## VIOLET RED BILE LACTOSE AGAR

CM0107

### Typical Formula\*

Yeast extract	grams per litre	3.0
Peptone		7.0
Sodium chloride		5.0
Bile salts No.3		1.5
Lactose		10.0
Neutral red		0.03
Crystal violet		0.002
Agar		12.0

\* adjusted as required to meet performance standards

### Directions

Suspend 38.5g in 1 litre of distilled water. With frequent agitation, bring to the boil to dissolve completely. Cool to 50°C. Mix well and pour into sterile Petri dishes or hold at 45°C when using the pour plate technique. DO NOT AUTOCLAVE.

### Physical Characteristics

Straw/pink, free-flowing powder  
 Colour on reconstitution - dark purple  
 Moisture level - less than 7%  
 pH 7.4 ± 0.2 at 25°C  
 Clarity - clear  
 Gel strength - firm, comparable to 12.0g/litre of agar

### Microbiological Tests Using Optimum Inoculum Dilution

Control Medium: Tryptone Soya Agar

### Reactions after incubation at 30 ± 2°C for 24 ± 2 hours

Inoculation using pour plate technique

Medium is challenged with 50-150 colony-forming units

<i>Klebsiella pneumoniae</i>	ATCC®29665	1-2mm purple/pink colonies, slight halo
<i>Proteus mirabilis</i>	ATCC®12453	Pinpoint-0.5mm purple/pink colonies, no halo

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A satisfactory result for pour plate technique is represented by recovery of positive strains equal to or greater than 50% of the control medium.

There shall be no gassing in the medium.

Inoculation using surface plate technique

Medium is challenged with 10-100 colony-forming units

<i>Shigella sonnei</i>	ATCC®25931	1-3mm straw colonies
<i>Enterobacter aerogenes</i>	ATCC®13048	0.5-2mm pink colonies, dark centre
<i>Pseudomonas aeruginosa</i>	ATCC®27853	1-5mm colourless/straw colonies

A satisfactory result is represented by recovery of positive strains equal to or greater than 50% of the control medium.

Medium is challenged with 1E+04 to 1E+06 colony-forming units

<i>Staphylococcus aureus</i>	ATCC®5923	No growth
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Negative strains are inhibited.

Medium is challenged with 1E+04 to 1E+06 colony-forming units

<i>Proteus mirabilis</i>	ATCC®12453	0.5-2mm straw colonies, no swarming
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## Testing performed in accordance with ISO11133:2014

### Reactions after incubation at 30 ± 2°C for 24 ± 2 hours


Inoculation using pour plate technique

Medium is challenged with 50-120 colony-forming units

<i>Escherichia coli</i>	ATCC®8739	WDCM00012	1-3mm purple/pink colonies and purple halo
<i>Escherichia coli</i>	ATCC®25922	WDCM00013	1-3mm purple/pink colonies and purple halo

A satisfactory result is represented by recovery of positive strains equal to or greater than 50% of the control medium.

There shall be no gassing in the medium.

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Medium is challenged with 1E+04 to 1E+06 colony-forming units

<i>Pseudomonas aeruginosa</i>	ATCC®27853	WDCM00025	1-5mm colourless/straw colonies
<i>Enterococcus faecalis</i>	ATCC®19433	WDCM00009	No growth
<i>Enterococcus faecalis</i>	ATCC®29212	WDCM00087	No growth

## Revision History

Section / Step	Description of Change	Reason for Change	Reference
All	Reformatting of document to new format	N/A	N/A
<i>Pseudomonas aeruginosa</i> ATCC® 27853	Change of colony size	QC specification change	BT-CC-2176