

# Pierce<sup>®</sup> NeutrAvidin<sup>®</sup> Coated 96-Well Plates

0611.4

Number	Description
15127	Pierce NeutrAvidin Coated Plates (clear, 8-well strips), 5 each
15129	Pierce NeutrAvidin Coated Plates (clear, 96-well), 5 each
15116	Pierce NeutrAvidin Coated Plates (white, 96-well), 5 each
15117	Pierce NeutrAvidin Coated Plates (black, 96-well), 5 each Activation Level: 100µL Binding Capacity: ~15pmol D-biotin/well Blocking Buffer: These plates are supplied blocked with SuperBlock <sup>®</sup> Blocking Buffer
15128	Pierce NeutrAvidin Coated Plates (clear, 8-well strips), 5 each
15123	Pierce NeutrAvidin Coated Plates (clear, 96-well), 5 each
15216	Pierce NeutrAvidin Coated Plates (white, 96-well), 5 each
15217	Pierce NeutrAvidin Coated Plates (black, 96-well), 5 each Activation Level: 200µL Binding Capacity: > 15pmol D-biotin/well Blocking Buffer: These plates are supplied blocked with Blocker <sup>™</sup> BSA

**Storage:** Upon receipt store plates at 4°C in unopened pouches. Once opened, place unused plates in a resealable bag with desiccant and store at 4°C. Plates are shipped at ambient temperature.

## Introduction

The Thermo Scientific Pierce NeutrAvidin Coated Plates are ideal for capturing biotin-labeled molecules without interference from nonspecific binding. NeutrAvidin Protein is deglycosylated avidin, which reduces lectin binding to undetectable levels while retaining stability and biotin-binding affinity. NeutrAvidin Protein offers the advantages of a near-neutral pI (6.3), to minimize nonspecific adsorption, and the lack of the RYD sequence, which eliminates nonspecific binding to the RGD binding domain of adhesion receptors present in a variety of cells. NeutrAvidin Protein yields the lowest nonspecific binding among the known biotin-binding proteins. The clear, white and black plates can be used with colorimetric, chemiluminescent and fluorescent detection methods, respectively.

## Example ELISA Protocol using NeutrAvidin Coated Plates

### A. Materials Required

- Wash Buffer: Tris-buffered saline (25mM Tris, 150mM NaCl; pH 7.2; Product No. 28376), 0.1% BSA, 0.05% Tween<sup>®</sup>-20 Detergent; alternatively, use Thermo Scientific Blocker BSA (Product No. 37520) supplemented with 0.05% Tween-20 Detergent
- Biotinylated capture antibody adjusted to 10µg/mL, or other appropriate concentration, with Wash Buffer
- Antigen adjusted to appropriate concentration with Wash Buffer
- Primary antibody adjusted to appropriate concentration with Wash Buffer
- Enzyme-labeled secondary antibody adjusted to appropriate concentration with Wash Buffer
- Appropriate enzyme substrate: example substrates are the Thermo Scientific TMB Substrate Kit (Product No. 34021) for horseradish peroxidase and the Phosphatase Substrate Kit (Product No. 37620) for alkaline phosphatase

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**B. Method**

1. Wash each well three times with 200µL of Wash Buffer. Add 100µL of the biotinylated capture antibody to each well and incubate for 2 hours at room temperature.
2. Wash each well three times with 200µL of Wash Buffer. Make a serial dilution of the antigen and add 100µL to each well. Incubate plate for 30 minutes at room temperature.
3. Wash each well three times with 200µL of Wash Buffer. Add 100µL of the primary antibody to each well and incubate plate for 30 minutes at room temperature.
4. Wash each well three times with 200µL of Wash Buffer. Add 100µL of the enzyme-labeled secondary antibody to each well. Incubate plate for 30 minutes at room temperature.
5. Wash each well three times with 200µL of Wash Buffer.
6. Follow the manufacturer's instructions for the specific detection system.

**Procedure for Determining Binding Activity of the NeutrAvidin Coated Plates**

The binding activity of the plates may be tested using Thermo Scientific Biotinylated Alkaline Phosphatase (Product No. 29339) and PNPP (Product No. 37620) or Biotinylated Horseradish Peroxidase (Product No. 29139) and TMB (Product No. 34021).

1. Rinse each well with three times with 200µL of wash buffer (e.g., TBS).
2. Prepare a 1mg/mL solution of the biotinylated enzyme. Make 1:2 serial dilutions using a 1:1000 dilution for the first well. Incubate the wells for 1 hour at room temperature.
3. Wash each well three times with 200µL of TBS containing 0.05% Tween-20 Detergent.
4. Incubate with 100µL of substrate solution for 15 minutes at room temperature.
5. Measure the absorbance of each well. Active plates result in an absorbance of 0.5 to 1.0 OD at 405nm.

**Related Thermo Scientific Products**

<b>37070</b>	<b>SuperSignal® ELISA Pico Chemiluminescent Substrate, 100mL</b>
<b>15169</b>	<b>QuantaBlu™ Fluorogenic Peroxidase Substrate Kit</b>
<b>34028</b>	<b>1-Step Ultra TMB-ELISA, 250mL</b>
<b>37621</b>	<b>1-Step PNPP, 100mL</b>
<b>29339</b>	<b>Biotinylated Alkaline Phosphatase, 1mg</b>
<b>29139</b>	<b>Biotinylated Horseradish Peroxidase, 5mg</b>
<b>15075</b>	<b>Reagent Reservoirs, 200/pkg</b>
<b>15082</b>	<b>Microtube Racked System, 960 tubes</b>
<b>15036</b>	<b>Sealing Tape for 96-Well Plates, 100/pkg</b>

**General References**

- Denlinger, L.C., *et al.* (2001). Cutting Edge: The nucleotide receptor P2X7 contains multiple protein- and lipid-interaction motifs including a potential binding site for bacterial lipopolysaccharide. *J Immunol* **167**:1871-6.
- Ferre-Aubineau, V., *et al.* (1995). Colorimetric microtiter plate hybridization assay using monoclonal antibody for detection of an amplified human immunodeficiency virus target. *J Virol Meth* **55**:145-51.
- Hiller, Y. *et al.* (1987). Biotin binding to avidin. Oligosaccharide side chain not required for ligand association. *Biochem J* **248**:167-71.
- Holmstrom, K., *et al.* (1993). A highly sensitive and fast non-radioactive method for detection of polymerase chain reaction products. *Anal Biochem* **209**:278-83.
- Simon, M.D., *et al.* (2004). A phage display selection of engrailed homeodomain mutants and the importance of residue Q50. *Nucl Acid Res* **32(12)**:3623-31.

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